Evaluation of Analgesic, Antidiarrheal and Anti-hyperglycemic Activities of *Dactyloctenium austral* (Poaceae)

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**Abstract**

*Dactyloctenium austral* belongs to the family of Poaceae. It is also called gramineae or true grasses. Poaceae is the fifth largest family of flowering plants. The current study was conducted on methanol extract of the aerial parts of *D. austral* (MEDA) to evaluate its in vivo analgesic activity by acetic acid-induced writhing method in mice. The plant extract was also evaluated for antidiarrheal and anti-hyperglycemic activities using castor oil-induced diarrhea and oral glucose tolerance test, respectively. In acetic acid-induced writhing test, the extract showed 52.18% and 62.40% inhibition of writhing at the doses of 200-400 mg/kg body weight, respectively while standard aspirin at the dose of 50 mg/kg bw showed 58.12% writhing inhibition. In anti-hyperglycemic test, the extract revealed its activity in a dose dependent manner. In antidiarrheal activity test, the extract exhibited 48.54% and 72.92% inhibition of defecation at the doses of 250-500 mg/kg bw, respectively whereas the standard loperamide (3 mg/kg bw) displayed 70.24% inhibition of defecation.

**Key words:** *Dactyloctenium austral*, analgesic, antidiarrheal, anti-hyperglycemic.

**Introduction**

A medicinal plant is any plant which contains substances that can be used for therapeutic purposes or which is a precursor for synthesis of useful drugs (Sofowora, 1982). Chemical principles from natural sources have become much simpler and have contributed significantly to the development of new drugs from medicinal plants (Meena et al., 2010). Traditional medicines are very important since early ancient period because of their faithfulness in the use against various ailments and human sufferings. Different types of bioactive natural compounds are derived from medicinal plants and they serve as raw materials for new drug discovery (Ramawat et al., 2009). In last few years, there has been great focus on the possible health benefits of natural substances with antioxidant, antimicrobial, analgesic, anticancer, antidiabetic and others activities. This has resulted in an enormous increase of research on different medicinal plants to find lead compounds responsible for such pharmacological activities.

*Dactyloctenium austral* is a 32-80 cm long evergreen creeping perennial grass species, which belongs to the family Poaceae (Gräser). The Poaceae (also called Gramineae, Durban Grass or true grasses) are a large family of flowering plants. Poaceae is the fifth-largest plant family, following the Orchidaceae, Asteraceae, Fabaceae, and Rubiaceae (Firth et al., 2002).

Among different disorders, diarrhea is one of the causes of morbidity and mortality especially in developing countries (Carlos and Saniel, 1990). The World Health Organization (WHO) has established a diarrhea disease control program for the treatment and management of diarrhea that includes traditional medicine practices along with health education and prevention approaches, which is mostly based on herbal products (WHO, 2015). WHO has accepted
traditional medicine as an alternative health care form. In a developing country like Bangladesh, where a handsome amount of people including children are affected by diarrhea every year, the search for plants with antidiarrheal activity that could be used against diarrheal disease is of prime interest.

Some antidiabetics have serious side effects and deleterious contraindications. Hence, researchers are paying attention to herbal medications having high therapeutic efficacy with minimal side effects. The antidiabetic agents from medicinal plants are very promising and traditionally acclaimed medicinal plants are being investigated for their antidiabetic potential (Babu et al., 2002; Parthasarathy et al., 2009). In other studies, the crude extracts of some medicinal plants were evaluated for alpha-amylase, alpha-glucosidase, total phenolic and total flavonoid contents where the extract have shown significant anti-hyperglycemic and analgesic activities (Das et al., 2012; Sangeetha et al., 2012; Telagari et al., 2015). In this context, we hypothesize that the leaf extracts of this plant might possess some compounds with anti-hyperglycemic, antidiarrheal and analgesic activities.

Materials and Methods

Collection of plant material: The aerial parts of *D. australe* were used as the raw material for the extraction and other investigation process. The plant sample was collected from Sylhet, Bangladesh in April, 2016. Any types of undesirable materials or plants or plant parts were separated from the collected plant parts. The plant was identified in Bangladesh National Herbarium, Dhaka, where a voucher specimen (Accession number: DACB-12771) has been deposited.

Drying and grinding: The collected plant materials were washed with water. After washing it was subjected to shed-drying for 1-2 weeks. When the plant parts were suitable for grinding, it was ground to a coarse powder by a grinder (Wuhu motor factory, China). Finally, the powder material was stored in a sealed container and kept in a dark, cool and dry place until farther processing.

Cold extraction: About 700 g of finely powered plant materials was taken in a clean glass container and it was soaked in 1500 mL of methanol. Then the container was sealed and kept for a period of 15 days. During this time, it was subjected to occasional stirring and shaking. The mixture was then filtered by cotton. Finally, it was filtered by Whatman filter paper number 1 and concentrated to give methanol extract of *D. australe* (MEDA).

Experimental animals: About 4-5 weeks aged Swiss-albino mice of both sexes (70 mice), average weight of 20-35g was collected from central animal house of the Department of Pharmacy, Jahangirnagar University, Savar, Dhaka-1342 and were used for the present study. The animals were randomly selected and divided into normal and experimental groups. After one week for their adaptation, all the experimental processes was conducted and these were performed in an isolated and noiseless environmental condition. The animals were housed under standard laboratory conditions (relative humidity 55-65%, room temperature 25.0 ± 20 °C, and 12 h light dark cycle) and fed with standard diet (icddr,b formulated) and had free access to tap water but were fasted 12 h prior to each experiment. The Federation of European Laboratory Animal Science Associations (FELASA) guidelines and recommendations were followed to reduce the pain and stress of the experimental mice.

Acetic acid-induced writhing test: The peripheral analgesic activity of the samples was evaluated in mice using acetic acid-induced writhing method (Koster et al., 1959; Biswas et al., 2009; Hossain et al., 2016; Woolfe et al., 1969). Mice were divided into 4 groups of 5 mice in each group. The control group received 1% Tween 80 in normal saline (10 ml/kg body weight), the standard group received Aspirin (50 mg/kg bw) and the experimental groups received crude extract of 200-400 mg/kg bw. Forty minutes later, each mouse was injected with 1% acetic acid at a dose of 10 ml/kg bw. The number of writhing responses was recorded for each animal during a subsequent 5 min period after 10 min of intraperitoneal administration of acetic acid and the mean writhing for each group was obtained.
The percentage inhibition was calculated using the formula –

\[
\text{% Inhibition} = \frac{\text{Mean number of writhing (control)} - \text{Mean number of writhing (drug)}}{\text{Mean number of writhing (control)}} \times 100
\]

*Evaluation of antidiarrheal activity:* To evaluate castor oil-induced antidiarrheal activity of the extract, the experimental mice were also divided into four groups: two test groups, control and standard consisting of 5 mice in each group. Tween-80 (1% in water) was given to the control group and the test groups of mice were given the extract at 200-400 mg/kg bw. Loperamide (3 mg/kg bw) was used as the standard drug. After 60 minutes, each mouse of all groups was administered 0.5 ml of castor oil in oral route. All animals were then kept separately in transparent cage having white blotting paper in order to count the number of faces. Every hour the blotting paper was changed and it was observed for a period of 4 hours. Latent period of fecal drops and percent inhibition of defecation of each group were determined (Rahman et al., 2010; Hasan et al., 2017; Islam et al., 2013). By using the following formula the percent inhibition of defecation was calculated:

\[
\text{Percent inhibition} = \frac{(D_0 - D_1)}{D_0} \times 100 \%
\]

Where, \(D_0\) is the number of defecation of the control group, and \(D_1\) is the number of defecation of the test or standard group.

*Evaluation of anti-hyperglycemic activity:* To evaluate the anti-hyperglycemic activity of the extract, the experimental mice were also divided into four groups: two test samples, control and standard consisting of 5 mice in each group. The test groups of mice were given the extract at 250-, 500- and 1000 mg/kg bw. Glibenclamide (5 mg/kg bw) was used as a standard drug. In a fasting state (having no food for at least 10 hours but not more than 16 hours), the experimental animals were tested. After selection and weighing of mice, fasting blood glucose level for control, standard and two test groups were measured. Then glucose solution was administered and blood glucose levels after 30, 60, and 150 min were measured (Kumar et al., 2006; Shirwaikar et al., 2006; Kessler et al., 2005). To estimate blood glucose level, blood samples of experimental mice were drawn by pricking with a sterile needle in the tail vein. The blood glucose levels were measured by using the glucometer and compatible blood glucose strips. The blood glucose levels were measured in millimole per liter (mmol/l) unit.

*Statistical analysis:* The values are presented as mean ± standard error of mean (SEM) and one-way ANOVA analysis was used to determine the significant difference between the control group and experimental groups, the p values < 0.05 were considered to be statistically significant.

**Results and Discussion**

*Acetic acid-induced writhing test:* The results of the test showed that MEDA at the dose of 200-400 mg/kg bw exhibit significant (p < 0.01) inhibition of writhing reflex by 52.18% and 62.40%, respectively while the standard (Aspirin, 50 mg/kg bw) drug was found to inhibit the writing response by 58.12% (Table 1).

*Evaluation of antidiarrheal activity:* In the castor oil-induced diarrhea, the MEDA at the doses of 200-400 mg/kg bw of mice significantly (p < 0.001) decreased the total number of faeces as well as delayed the onset of diarrhea in a dose dependent manner. Percent inhibition of defecation at doses 200-400 mg/kg bw was 48.54 and 72.92, respectively whereas that for the standard loperamide (3 mg/kg) was 70.24 (Table 2).

*Evaluation of anti-hyperglycemic activity:* The extract of *D. australis* showed significant (p< 0.02) anti-hyperglycemic activity on mice in a time dependent manner as compared to the control groups (Table 3).
Plants are natural sources of diverse therapeutic activities (Raja et al., 2013). Accordingly, the crude drugs are getting much more acceptances in healthcare sector (Joshi et al., 2011). In pathological conditions, tissue injury causes pain resulting in the local release of chemical mediators such as prostaglandins, cytokinins, leukotrienes etc. They act on the nerve terminals in both activating them directly and enhancing their sensitivity to other stimulation (Kanodia et al., 2008; Goldstein et al., 1970). Acetic acid administration through intraperitoneal routes can produce pain by consequent abdominal writhing due to the release of mediators like prostaglandin E₂ and other lipooxygenase products (Sulaiman et al., 2008). Prostaglandin mainly prostacyclines (PGI₂) and prostaglandin E (PG-E) are responsible for pain sensation due to the excitation of δ-nerve fibers (Lourens et al., 2004; Islam et al., 2015). Thus, the extract of *D. austral* may produce non-narcotic

### Table 1. Analgesic activity of methanol extract of *D. austral* leaves in acetic acid-induced pain in mice.

<table>
<thead>
<tr>
<th>Treatment (n=5)</th>
<th>Dose (mg/kg)</th>
<th>No of writhes</th>
<th>% Inhibition</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>1% Tween-80</td>
<td>40.83 ± 1.13</td>
<td>---</td>
</tr>
<tr>
<td>Aspirin</td>
<td>50</td>
<td>17.43 ± 3.05*</td>
<td>58.12</td>
</tr>
<tr>
<td>MEDA</td>
<td>200</td>
<td>19.93 ± 0.33*</td>
<td>52.18</td>
</tr>
<tr>
<td>MEDA</td>
<td>400</td>
<td>15.35 ± 0.55*</td>
<td>62.40</td>
</tr>
</tbody>
</table>

Values are expressed as mean ± S.E.M (n = 5). *p < 0.01, compared with vehicle control (ANOVA followed by Dunnet’s t-test).

### Table 2. Effects of methanol extract of *D. austral* leaves on castor oil-induced diarrhea in mice.

<table>
<thead>
<tr>
<th>Group</th>
<th>n</th>
<th>Onset of diarrhea (mean ± SEM, min)</th>
<th>Number of stools after 4 hrs (mean± SEM)</th>
<th>Inhibition of defecation (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control (1% tween-80 water)</td>
<td>5</td>
<td>33 ± 3.59</td>
<td>12.3 ± 0.88</td>
<td>70.24</td>
</tr>
<tr>
<td>Standard (Loperamide 3 mg/kg)</td>
<td>5</td>
<td>197.4 ± 2.65*</td>
<td>3.66 ± 0.0014*</td>
<td></td>
</tr>
<tr>
<td>MEDA 200 mg/kg bw</td>
<td>5</td>
<td>101 ± 3.64*</td>
<td>6.33 ± 0.023*</td>
<td>48.54</td>
</tr>
<tr>
<td>MEDA 400 mg/kg bw</td>
<td>5</td>
<td>178.8 ± 5.0*</td>
<td>3.33 ± 0.74*</td>
<td>72.92</td>
</tr>
</tbody>
</table>

* p <0.001 vs control group. SEM: standard error of mean. n = number of mice

### Table 3. Effect of the methanol extract of *D. austral* leaves on oral glucose tolerance test (anti-hyperglycemic activity test) in normal control mice.

<table>
<thead>
<tr>
<th>Group (n = 5)</th>
<th>Blood glucose level (mean ± SEM)</th>
<th>0 min</th>
<th>30 min</th>
<th>60 min</th>
<th>120 min</th>
<th>180 min</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td></td>
<td>6.2 ± 0.4</td>
<td>10.5 ± 0.3</td>
<td>7.3 ± 0.6</td>
<td>5.7 ± 0.1</td>
<td>7.05 ± 0.55</td>
</tr>
<tr>
<td>Glibenclamide 5mg/kg*</td>
<td></td>
<td>5.45 ± 0.25</td>
<td>4.8 ± 0.1</td>
<td>3.5 ± 0.2</td>
<td>1.7 ± 0.2</td>
<td>6 ± 4</td>
</tr>
<tr>
<td>MEDA 250 mg/kg**</td>
<td></td>
<td>4.05 ± 0.75</td>
<td>5 ± 1.9</td>
<td>5.55 ± 0.55</td>
<td>4.7 ± 1.6</td>
<td>4.2 ± 0.0</td>
</tr>
<tr>
<td>MEDA 500 mg/kg***</td>
<td></td>
<td>3.6 ± 0.8</td>
<td>4.8 ± 1.2</td>
<td>4.6 ± 0.6</td>
<td>4 ± 1.7</td>
<td>3.65 ± 0.45</td>
</tr>
<tr>
<td>MEDA 1000 mg/kg****</td>
<td></td>
<td>5.85 ± 0.65</td>
<td>4.45 ± 1.45</td>
<td>4.15 ± 0.15</td>
<td>5.55 ± 0.45</td>
<td>4.04 ± 0.25</td>
</tr>
</tbody>
</table>

Here, *p = 0.0276, **p = 0.0169, ***p = 0.2868, ****p = 0.6014 vs Control group. SEM: standard error of mean.
analgesic activity due to the inhibition of prostaglandin synthesis by blocking of lipoxygenase and cyclooxygenase activities. In the present study, the methanolic extract of *D. australe* showed significant (p < 0.01) writhing inhibitions compared to that of standard aspirin. The result also showed that percentage of writhing inhibition in two test samples (250-500 mg/kg bw) are sufficient enough to produce analgesic activity as compared to standard test results (Table 1). It is clear that the standard drug aspirin is a potent analgesic and it can produce stronger analgesic activity rather the methanolic extract of *D. australe*.

Several studies have shown that many medicinal plants have shown bioactivities, for instance, delay or suppress gut motility, intestinal transit, stimulate water absorption or reduce intraluminal fluid accumulation (Gutierrez et al., 2013). Diarrhea refers to excess passage of watery stools which is caused by decreased consistency or increased frequency of bowel movements. The possible mechanism of diarrhea would be the change in active ion transport system either by increasing chloride secretion or decreasing sodium absorption. It may also include the increase in luminal osmolality, change in intestinal motility and increase in tissue hydrostatic pressure (Schiller, 1995). Among all the mechanisms, castor oil (active compound ricinoleic acid) induces diarrhea by stimulating intestinal motility and secretory processes, is the major one (Neimegeers et al., 1984). In the present study, anti-diarrheal activity of the plant extract of *D. australe* was investigated by castor oil-induced diarrhea in mice. The result showed that MEDA significantly (p<0.001) expressed anti-diarrheal activity in a dose dependent manner that makes it more similar with other plant extracts (Table 2) (Umer et al., 2013).

Diabetes is a metabolic disorder that indicates elevated blood glucose concentration and occurred by insufficient insulin secretion and action (Henriksen, 2001; Schroeder and Koltermann, 2010). The agents that are used to treat diabetes by means of decreasing blood glucose concentration or sufficiently secreting insulin are known as anti-hyperglycemic agents. As per reference of Joy and Kuttan, (1999) the mechanism of anti-hyperglycemic agents had either by potentiating pancreatic insulin secretion or increasing glucose uptake. Such mechanism has also been proposed in another study, root extracts of *Helicteres isora* (Venkatesh et al., 2004). The current study with methanolic extract of *D. australe* expressed significant (p < 0.001) anti-hyperglycemic activity as compared to control and other groups (Table 3). The possible mechanism of this activity may be either by potentiating pancreatic insulin secretion or increasing glucose uptake. The results also demonstrate that animals treated with standard drug (Glebenclamide 5 mg/kg bw) has shown strong anti-hyperglycemic activity rather than the other groups and the two test groups (250-500 mg/kg bw) also revealed significant decrease in blood glucose level. From the above discussion, we can say, that the MEDA may be a potential source of anti-hyperglycemic agents.

**Conclusion**

The present study summarizes analgesic, antidiarrheal and anti-hyperglycemic activities of *D. australe* leaf extract. However, further studies are necessary to explain the probable mechanisms related with these bioactivities.

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**References**


